

Morgan Olmstead

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RESEARCH INTERESTS

Genetic engineering, agricultural pests, insects as feed and food, insect microbiomes, functional genomics, insect behavior, and sequencing technologies.

EDUCATION

North Carolina State University, Raleigh, NC **Aug 2023 – Present**

Ph.D. in Entomology, Genetic Pest Control Focus

Advisor: Dr. Marcé Lorenzen

Genetics and Genomics Fellowship

Interdisciplinary Perspectives on Genes and Genomes Graduate Minor (IPGG)

Genetic Engineering and Society (GES) Interdisciplinary Minor

University of New Hampshire, Durham, NH **Aug 2016 – Jun 2018**

M.S. in Integrative and Organismal Biology

Advisor: Dr. Carrie Hall

Thesis: “The microbiome of four North American *Nicrophorus* species (Coleoptera: Silphidae): Implications for parental care behavior in insects”

Ohio University, Athens, OH **Aug 2012 – May 2016**

B.S. in Biological Sciences

Minor in Anthropology

Certificate in Global Health

RESEARCH AND TEACHING EXPERIENCE

North Carolina State University, Raleigh, NC **Aug 2023 – Present**

Graduate Student, Dr. Marcé Lorenzen, Entomology

- Exploring selfish genetic elements in *Tribolium castaneum* agricultural pest
- Engineering yellow mealworm (*Tenebrio molitor*) for traits in the Insects as Food and Feed industry

U.S. Department of Agriculture – ARS, CGAHR, Manhattan, KS **Nov 2020 – Jun 2023**

Research Technician, Stored Product Insect and Engineering Research

- Generated long-read sequencing data with Oxford Nanopore technology; transcriptome and functional genomics sequencing and variant analysis studies on yellow mealworm strains (*Tenebrio molitor*); shotgun metagenomic and pathogen studies in yellow mealworms, naval orange worm (*Amyelois transitella*), and Diptera; various extraction and sequencing kits and protocols
- Manually annotated insect genes (*Rhyzopertha dominica*)
- Ran behavioral assays with *T. molitor* larvae infected with *Bt* toxin, bioassays of *Tribolium castaneum* reproduction on alternative diets, and bioassays of mites commonly found in stored grain
- Developed rearing protocols for superworm (*Zophobas morio*), lesser mealworm (*Alphitobius diaperinus*), and banded crickets (*Gryllodes sigallatus*); and maintained rearing of yellow mealworms (*Tenebrio molitor*), Indian meal moths (*Plodia interpunctella*), red flour beetles (*T. castaneum*), rice weevils (*Sitophilus oryzae*), and lesser grain borer (*R. dominica*)
- Collaborated with other departments within the USDA, APHIS, FDA, and USAID as well as companies and universities on projects involving using Insects as Food and Feed
- Performed lab management duties such as purchasing and keeping track of accounting with purchase card, sat on a hiring committee, and mentored undergraduate and graduate students

Vanderbilt University Medical Center, Nashville, TN **Aug 2018 - Aug 2020**
Research Assistant II, VANTAGE sequencing core

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- Extracted DNA and RNA from FFPE samples utilizing Covaris extraction methods
- QC of DNA and preparation of genomic and RNA libraries such as WGS, exomes, mRNA and rRNA depletion transcriptome libraries, and single cell sequencing using 10x
- Prepared and loading of Illumina sequencing instruments, utilized BluePippin for size selection
- Trained lab members on library preparation, QC instrumentation, and sequencer loading

Vanderbilt University, Nashville, TN

Volunteer, Dr. Ann Tate, Dept Biological Sciences

Oct 2019 - Feb 2020

- Conducted metabolism tests using respirometry techniques on *T. castaneum*

University of New Hampshire

Aug 2016 - Jul 2018

Graduate Student, Dr. Carrie Hall, Dept Biological Sciences

- Researched the effect of microbial symbionts on insect reproductive behavior of *Nicrophorus marginatus* (Coleoptera: Silphidae) and classified microbial communities of five *Nicrophorus* spp.
- Developed a dissection protocol to identify and remove tissues from beetle specimens, including reproductive tissues of both males and females, salivary glands, and digestive tissues
- Extracted and purified DNA from microorganisms found within beetle tissue
- Analyzed 16s and ITS microbiome data using the software program QIIME 2 and supplementally in R
- Studied the effects of microbial communities on parental care behavior and offspring development through the administration of antibiotics and analyzed the behavioral metrics in R

University of New Hampshire Biology Department, Durham, NH

Aug 2016 - May 2018

Teaching Assistant, Introduction to Biology, Evolution, Biodiversity, and Ecology; Forensic Biology

- Taught methods used in ecology research, including collection and identification of animals and plants, research design, writing reports, and basic statistics utilizing excel
- Graded lab reports and homework, helped prepare lessons and quizzes for the lab course

Center of Science and Industry, Columbus, OH

Jun 2015 - Nov 2015

Experience Programs Teacher

- Performed scientific experiments for guests and guided guests through their own experiments

Ohio University

Fall 2015

Volunteer, Dr. Kelly Johnson, Dept of Biological Sciences

- Collected and identified aquatic insects at family level

Ohio University Anthropology Department, Athens, OH

Aug 2014 - Dec 2014

Peer-Led Team Learning Teacher, Introduction to Biological Anthropology

- Created supplementary material for students in lab including class activities
- Managed and executed exam review sessions

PUBLICATIONS

1. **Olmstead, M.L.**, VanDonge, K.N., Swistek, S.E., Shultz, P.T., Cohnstaedt, L.W., Oppert, B.S. (In Prep). Microbiome and pathogens of filth flies collected from Chicken and Dairy farms for use as protein supplement in animal feed.
2. VanDonge, K.N., Swistek, S.E., **Olmstead, M.L.**, Mota-Peynado, A., Ewing, R.D., Mitzel, D., Oppert, B.S., Cohnstaedt, L.W., Shults, P.T. (2024). Safety of wild-caught *Musca domestica* for use as protein supplement in animal feed. *Journal of Economic Entomology*. <https://academic.oup.com/jee/advance-article/doi/10.1093/jee/toad239/7517006>
3. Oppert, B. et al. (2023). The Genome of the Yellow Mealworm, *Tenebrio molitor*: it's Bigger Than You Think. *Genes* 14(12). <https://www.mdpi.com/2073-4425/14/12/2209>
4. Leierer, D., **Olmstead, M.**, Oppert, B. (2023). Sequencing to identify pathogens in *Tenebrio molitor*.

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1-740-317-6401

Implications in insects farmed for food and feed. *Frontiers in Insect Science* 3.

<https://www.frontiersin.org/articles/10.3389/finsc.2023.1059046/full>

5. Oppert, B. et al. (2022). The Genome of *Rhyzopertha dominica* (Fab.) (Coleoptera: Bostrichidae): Adaptation for Success. *Genes* 13(3), 446. <https://www.mdpi.com/2073-4425/13/3/446>

CONFERENCE PRESENTATIONS

Oral presentation

- 1. BASF Nora Spring Review** **Mar 2024**
Morgan Olmstead
"CYP450 Gene Expression in Honeybees (*Apis mellifera*)"
- 2. Plant and Animal Genome Conference** **Jan 2023**
Application of New Genomic Tools and Techniques in Arthropods workshop (Invited talk)
Morgan Olmstead, Brenda Oppert, Jose Morales-Ramos, Guadalupe Rojas, and Marcé Lorenzen
"Building a Bigger Beetles for Insects as Food and Feed"
- 3. Entomological Society of America Conference** **Oct 2021**
Advancing Insect Genomics through the Ag100Pest and i5K initiative symposium
Morgan Olmstead, Brenda Oppert
"Developing genetic resources for insect livestock in food application"
- 4. Ohio University Global Health Certificate Presentation** **Spring 2016**
Morgan Olmstead
"Pregnancy Traditions of Lhotshampa Refugee Population"

Poster presentation

- 1. Entomological Society of America Conference** **Oct 2021**
Morgan Olmstead, Dewey Leierer, and Brenda Oppert
"Utilization of a behavioral monitoring device to analyze movement and predict mortality in *Tenebrio molitor* exposed to *Bacillus thuringiensis* toxin"
- 2. Graduate Research Conference, University of New Hampshire** **Apr 2018**
Morgan Olmstead, Jeff Foster, and Carrie Hall
"Characterization of burying beetle (*Nicrophorus* spp.) microbiomes: Insights into the ecology and evolution of four North American species"
- 3. Entomological Society of America Conference** **Nov 2017**
Morgan Olmstead, Jeff Foster, and Carrie Hall
"Evidence for an evolutionary arms race: Behavioral compensation for endosymbiotic sex-ratio manipulation"

PROFESSIONAL DEVELOPMENT

Leadership

Leader of Entomology Graduate Program Peer Review Group, KSU	Spring 2023
Diversity, Equity, and Inclusion member with USDA	2022 - 2023
Secretary of Anthropology Club, Ohio University	2014 - 2015

Professional Affiliations

Entomological Society of America (member)	2017 - Pres
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Peer Review Manuscripts

(2) manuscript reviews for journal, PeerJ	Various
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Training and Workshops

ConGen 2022	Sep 2022
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2-week virtual workshop on the conceptual and practical aspects of population genetics, conservation genetics, and molecular ecology presented by University of Montana.

VectorBase Workshop

Jun 2022

Hands-on training for VectorBase databases and utilizing RNA-seq data from published datasets; held during Arthropod Genomics Symposium at Notre Dame

Vanderbilt University Microbiome Boot Camp

Nov 2019

Workshop on microbiome experimental design and hands-on learning with QIIME 2

New England Biolabs Molecular Biology Workshop

Jul - Aug 2018

Two weeks-long molecular biology workshop held at Smith College

MEDIA APPEARANCES/INTERVIEWS

Ohio University Arts and Sciences Alumni Spotlight

Aug 2017

"Happy Beginnings | Alum Enjoys Beetles Research in Grad School"

AWARDS AND GRANTS

Genetic Engineering Society Minor Fellowship (~17k)

Fall 2024

Genetics and Genomics Scholar Fellowship, NCSU (~32k)

2023-2024

USDA Employee Individual Special Acts or Service Awards (5)

Various

Sigma Xi, Grants-in-Aid of Research (\$937), UNH

May 2017

Travel Award, UNH

Fall 2017

Ohio University Gateway Scholarship

2012-2016

College of Arts and Sciences Scholar, OU

2012

OUTREACH AND MENTORSHIP

Mentorship

Molly Edeburn, undergrad at KSU

Jun 2022 – Jun 2023

Teaching skills in entomological bioassays, insect rearing, molecular benchtop, and experimental design.

Dewey Leierer, undergrad/PhD student at KSU

May 2021 – Jun 2023

Continued mentorship of undergraduate into PhD status including training in genomic techniques, writing, paper submissions, and several presentations.

Kortnee VanDonge, Veterinary Scholar Kansas State University

Summer 2022

Taught DNA extraction techniques and QC, experimental design techniques. Ended in poster presentation and eventual publication of results.

Sabrina Swistek, Veterinary Scholar student at Mississippi State

Summer 2022

Taught DNA extraction techniques and QC, experimental design techniques. Ended in poster presentation.

Adam Wallenfang, Biological Research Technician at USDA

Spring 2022

Taught mRNA library preparation using Lexogen kit, qPCR of Illumina libraries for sequencing.

Abby Perkins, undergrad at KSU

Oct 2021 - May 2022

Taught experimental design and reading of entomological bioassays. Ended in poster presentation.

Valerie Nguyen, master's student at KSU

Fall 2021

Advice and training on Illumina amplicon sequencing and analysis. Ended in poster presentation.

Outreach and volunteering

Science Olympiad hosted at NCSU

Jan 2024

Graded tests

FFA Insect Spectacular Event with USDA

Jun 2023

Gave children and adults tours of the USDA facility

K-State Open House

Apr 2023

Demonstrated USDA research on mealworms and Styrofoam for Manhattan, KS community

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Careers in government panel for KSU graduate students

Apr 2023

Member on panel for graduate students to ask questions about working for the government

Dover Middle School STEAM Academy

Feb 2017 - Apr 2017

Aided Ph.D. student in creating activities for middle schoolers in experimentation relating to the scientific method, utilizing insects and bacterial colonies

Historic Fort Steuben and Welcome Center

Summer 2013

Tour guide for the historic fort and archaeology dig in Steubenville, OH